

zic5 gene. We have generated in situ probes that are specific for the *zic1*, *zic2*, *zic3*, and *zic4* genes in chick. While there is considerable overlap in the expression patterns of these genes, there are also distinct differences. *zic2* and *zic3* are strongly expressed in the entire dorsal neural tube, including the brain, trunk, and tail tip. *zic1* expression is strong in the dorsal brain, but weaker in the dorsal neural tube of the trunk. *zic4* appears to be expressed exclusively in the head region. During somite development, *zic1* is expressed dorsomedially in epithelial somites, while *zic2* expression begins well after somites have given rise to distinct dermomyotome and sclerotome regions. *zic1–3* are expressed strongly in the dorsomedial parts of more mature somites, particularly in the posterior portions. *zic2* is the only family member expressed in the periotic mesoderm in the hindbrain. *zic2–4* are expressed in the developing eye and all *zic* genes are expressed in the optic stalk. Among the *zic* genes, only *zic2* is expressed in the limb buds. We are currently analyzing *zic3* and *zic4* expression in more detail.

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Program/Abstract # 172

A microarray screen for direct targets of the *Zic1* transcription factor

Sabah M. Hassan, Shuzhao Li, Christa Merzdorf
Department of Cell Biology and Neuroscience, Montana State University, Bozeman, MT, USA

The transcription factor *zic1* plays important roles in a variety of developmental processes. In vertebrates, these include patterning the early neural plate, development of the neural crest, somite development, and formation of the cerebellum. In addition, *zic1* promotes cell proliferation. To increase our understanding of the molecular mechanisms that underlie these processes, we have conducted a DNA microarray screen with Affymetrix gene chips to identify downstream target genes of *zic1*. The screen was performed using *Xenopus* ectodermal explants. By using an inducible *zic1* construct (*zic1GR*) and the protein synthesis inhibitor cycloheximide, the screen was designed to discover direct targets of the *Zic1* transcription factor. One new gene that was identified in this screen is the putative protease *Xfeb* that contributes to hindbrain patterning. We give a general overview of the genes identified in this screen and will discuss our findings from the screen with respect to genes involved in neural crest development and in anterior–posterior patterning.

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Program/Abstract # 173

Requirement of *Goosecoid* in early *Xenopus* development: A loss-of-function study

Veronika Sander, Bruno Reversade, E.M. De Robertis
Howard Hughes Medical Institute, University of California, Los Angeles, CA, USA

Goosecoid (*Gsc*), a homeobox gene, was the first gene found to be specifically expressed in Spemann's organizer. In the present study, we show that knock-down of *Gsc* in *Xenopus* by Morpholino oligomers (MO) leads to loss of head structures and expansion of ventral tissues. Unlike in the mouse, our results demonstrate a requirement of *Gsc* for head formation and axis patterning in early *Xenopus* development. In search of downstream mediator genes, we found that the effects of *Gsc* completely depend on the BMP antagonist Chordin. Ectopic expression of mouse *Gsc* mRNA lead to induction of a secondary axis. Co-injection of Chordin MO prevented this inductive capability of *Gsc* mRNA in 97% of the embryos. Evidence for regulation of Chordin by *Gsc* is further provided by quantitative PCR and in situ hybridization, showing that *Gsc* MO decreases *Chordin* expression in gastrulating embryos. Moreover, loss-of-function experiments in animal cap explants revealed mutual repression of *Gsc* and the ventral homeobox transcription factors *Vent1* and *Vent2*. MO-mediated knock-down of *Vent1/2* caused strong dorsalization of the embryos, which could be suppressed by *Gsc* MO. Our results suggest that *Gsc* and *Vent1/2* have opposing activities during gastrulation and act as regulators of each other in mesoderm patterning.

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Program/Abstract # 174

TFIIF trafficking and its nuclear assembly during early *Drosophila* embryo development

Javier Aguilar-Fuentes, Viviana Valadez-Graham, Enrique Reynaud, Mario Zurita
Department of Developmental Genetics and Molecular Physiology, Institute of Biotechnology, UNAM, Cuernavaca, Morelos, Mexico

We present the analysis of the dynamic of the transcription factor IIF (TFIIF) during early *Drosophila* embryo development. TFIIF consists of ten sub-units: cyclin-dependent kinase 7 (Cdk7), cyclin H and MAT1 (CAK), two helicases *Xpb/Hay* and *Xpd*, and p34, p44, p52, p42, p62 and p8 (core). We found that the TFIIF core is initially located in the cytoplasm of syncytial blastoderm embryos, after the mitotic division 10 and until the cellular blastoderm the core moves from the cytoplasm to the nucleus. The CAK complex has a different behaviour in early embryonic stages. The CAK is mostly cytoplasmic during cellularization and even during gastrulation. However, both components are positioned at promoters of genes that are activated at the onset of transcription. Later in development, the CAK complex becomes mostly nuclear and co-localizes in most chromosomal regions with the TFIIF core, but not in all sites, suggesting that the CAK complex could have a role in transcription of some loci without interaction with the TFIIF core. We also demonstrate that the CAK and the core coexist in the early embryo cytoplasm, however they do not interact until they are in the nucleus. Our data suggests that the complete assemble of the ten sub-units TFIIF occurs during the activation of zygotic transcription. In addition, we demonstrate that the